



Muse[®] Human CD4 T Cell Kit User's Guide

For Research Use Only. Not for use in diagnostic procedures.
4600-3423, Rev D
Catalog No. MIM100101 (100 tests)
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Application

The Muse® Human CD4 T Cell Kit is used in laboratory research studies to determine the CD4 T-cell count, total lymphocyte count, and CD4 T-cell percent of lymphocytes in human whole blood and peripheral blood mononuclear cell (PBMC) samples. Characterizing CD4 T-cell counts and percentages is important in evaluating immune status, understanding disease mechanism, in vaccine research and immunotoxicity studies. Minimal sample preparation is required in this no-wash assay to obtain accurate and precise results. The software provides:

- CD4 T-cell concentration in cells/ μ L
- CD4 T-cell percent of lymphocytes
- Total lymphocyte concentration in cells/ μ L

The Muse Human CD4 T Cell Assay is for use with the Guava® Muse Cell Analyzer. The Muse System makes sophisticated fluorescent-based analysis fast, easy, convenient, and affordable. Sample preparation is minimal, and after loading samples onto the Guava Muse Cell Analyzer, the intuitive software provides detailed or summary analysis of your cell sample in a few short steps.

Sufficient reagent is provided for the preparation and analysis of 100 tests.



CAUTION: This kit should not be used for patient diagnosis or patient management.

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Test Principle

CD4 T lymphocytes (T helper cells) are an important subpopulation of lymphocytes and play a central role in immune response. They stimulate B cells to secrete antibodies, recruit neutrophils to sites of infection, and secrete cytokines and chemokines to orchestrate immune response.

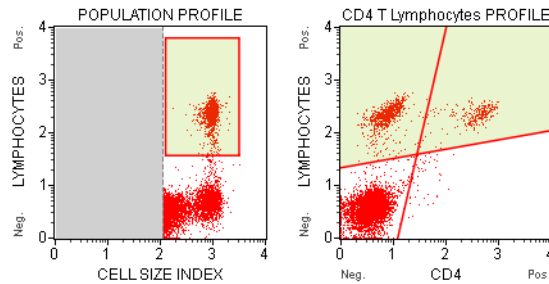
Normal T helper cells (defined with the use of an anti-lymphocyte cocktail and CD4 antibody¹) make up between 32% and 68% of the lymphocytes in healthy adults and show alteration of percent levels with age and development.² Multiple studies have determined the identification and enumeration of CD4 T-helper cells to be important in immunodeficiency, immunosuppression, and autoimmune diseases.³⁻⁶ Changes in T-cell populations have also been implicated in research on chronic inflammation associated with disease states, such as cancer and atherosclerosis, as well as in viral infections, bacterial infections, parasitic infections, sepsis, tuberculosis, burns, trauma, hepatitis, malnutrition, and stress.

The Muse® Human CD4 T Cell Kit allows for the detection and identification of lymphocytes and CD4 T lymphocytes in either whole blood or PBMCs using a simplified no-wash assay. The CD4 antibody identifies human helper/inducer CD4+ T cell (HLA Class II reactive) and recognizes a 60,000 Da surface antigen. Monocytes also express CD4 but at a lower density, and have no co-expression of the other antibodies present in the anti-lymphocyte cocktail; hence can be distinguished from CD4 T cells in this kit. The simplified identification can be of great utility in understanding the role of CD4 T lymphocytes in development, response, and mechanism of disease.

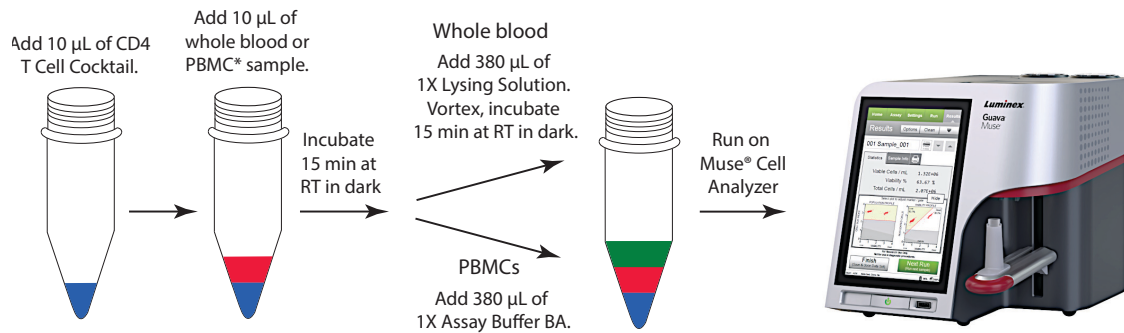
The Muse CD4 T Cell Assay uses an anti-lymphocyte cocktail that identifies the lymphocyte population, and CD4 antibody that binds to the CD4 cells. The assay provides results for CD4 T-cell lymphocytes, CD4 percent of lymphocytes, and the total lymphocyte count. Results in each of the four quadrants are indicated.

- UL: Non-CD4 Lymphocytes [Lymphocyte(+) and CD4(-)]
- UR: CD4 T Lymphocytes [Lymphocyte(+) and CD4(+)]
- LL: Negative Cells [Lymphocyte(-) and CD4(-)]
- LR: CD4 Non-Lymphocytes [Lymphocyte(-) and CD4(+)]

Figure 1: Representative plots from the Muse CD4 T Cell Assay. The first plot has a threshold marker, allowing you to eliminate debris based on cell size, as well as a gate to include lymphocytes. The second plot shows quadrant markers providing data on CD4 T lymphocytes.



Summary of Protocol



* PBMC samples should be in 1X Assay Buffer BA at a concentration of 4×10^6 to 2×10^7 cells/mL.

Kit Components

- Muse® Human CD4 T Cell Cocktail (Part No. 4700-1615, 100 tests/vial)
- Human 1X Lysing Solution (Part No. 4700-1620, 100 tests/vial)
- 1X Assay Buffer BA (Part No. 4700-1360, 100 tests/vial)

Materials Required but Not Supplied

- Guava® Muse® Cell Analyzer
- Whole blood or PBMC samples
- Micropipettors
- Disposable micropipettor tips
- Microcentrifuge tubes with screw caps, 1.5 mL (VWR, Catalog No. 16466-030, or equivalent)
- Vortex mixer
- Disposable gloves
- 100% bleach solution
- Deionized water
- Muse System Check Kit (Catalog No. MCH100101)
- Muse Count & Viability Reagent (Catalog No. MCH100102)
- Guava Fixative (Catalog No. 4700-0140), optional
- Guava ICF Instrument Cleaning Fluid (Catalog No. 4200-0140), optional

Precautions

- The Muse® Human CD4 T Cell Kit is intended for research use only; not for use in diagnostic procedures. This kit should not be used for patient diagnosis or patient management.
- Wear proper laboratory attire (lab coat, gloves, safety glasses) when handling or using this product.
- The 1X Assay Buffer BA and Muse CD4 T Cell Cocktail contains sodium azide, which is toxic if ingested. Reagents containing sodium azide should be considered a poison. If products containing sodium azide are swallowed, seek medical advice immediately and show product container or label. (Refer to NIOSH, National Institute for Occupational Safety and Health; CAS#: 2628-22-8; and also to GHS, The Globally Harmonized System of Classification and Labeling of Chemicals.) Aqueous solutions of sodium azide, when mixed with acids, may liberate toxic gas. Any reagents containing sodium azide should be evaluated for proper disposal. Sodium azide may react with lead and copper plumbing to form highly explosive metal azides. Upon disposal, flush with large volumes of water to prevent build-up in plumbing. Check with regulatory agencies to determine at what concentration sodium azide may cause a product to be regulated as hazardous.
- The instructions provided have been designed to optimize the kit's performance. Deviation from the kit's instructions may result in suboptimal performance and may produce inaccurate data.
- Some assay components included in the kit may be harmful. Please refer to the Safety Data Sheet (SDS) for specific information on hazardous materials.
- All fluorochrome conjugated antibodies are light-sensitive and must be stored in the dark at 2 to 8°C.
- During storage and shipment, small volumes of product may become entrapped in the seal of the product vial. For maximum recovery of product, centrifuge vial briefly prior to removing the cap.
- Avoid microbial contamination of the reagents, which may cause erroneous results.

- All biological specimens and materials should be handled as if capable of transmitting infection and disposed of with proper precautions in accordance with federal, state, and local regulations. Never pipette by mouth. Avoid specimen contact with skin and mucous membranes.
- Exercise care to avoid cross contamination of samples during all steps of this procedure, as this may lead to erroneous results.
- Do not use the reagent beyond the expiration date.
- Safety Data Sheets (SDSs) for kit reagents are available from our website (www.luminexcorp.com) or by contacting Luminex Technical Support.

Storage

- Store the Human CD4 T Cell Cocktail and 1X Assay Buffer BA at 2 to 8°C. Protect the CD4 T Cell Cocktail from exposure to light.
- Store the Human 1X Lysing Solution at room temperature (15 to 30°C).



CAUTION: Fluorochrome-conjugated antibodies should always be stored at 2 to 8°C. Any deviation in temperature for long periods of time may compromise the performance of the antibodies.

Before You Begin

Acquire samples shortly after the sample preparation is complete. While some donors have been shown to yield stable results for up to 4 hours, the stability of individual donors may vary. This time variability is a consequence of using live, unfixed cells. You should determine the stability results for your own samples. If longer stability is necessary, Guava Fixative may be added to the 1X Human Lysing Solution or the 1X Assay Buffer BA, as outlined in “Staining Protocol” on page 5.

This protocol was developed to allow direct enumeration of CD4 T cells in whole blood and PBMCs in cultures. Optimal staining is obtained for cell concentrations between 4×10^5 to 2×10^7 cells/mL. Luminex recommends using the Muse® Count & Viability Reagent to obtain accurate cell counts. Care should be taken to keep cell concentrations as constant as possible in all samples of an experiment.

Time considerations: The process of staining cells with the Muse Human CD4 T Cell Kit takes approximately 30 minutes for whole blood samples and 15 minutes for PBMCs. Acquiring data on the Muse Cell Analyzer takes approximately 2 minutes per sample.

Always run a System Check prior to performing the assay. For details, refer to the *Guava Muse Cell Analyzer User's Guide*.

Specimen Collection

Collect blood by venipuncture into a sterile K₃ EDTA (lavender top) or sodium heparin (green top) blood collection tube.



CAUTION: Discard blood samples that are hemolyzed, clotted, lipemic, discolored, or containing interfering substances.

Stain blood within 30 hours of collection for optimal results. Unstained anti-coagulated blood should be maintained at 18 to 25°C prior to sample processing.

Leave the capped tube of blood standing upright or lying on its sides if it is stored overnight. Do not rock or agitate blood in any way during extended storage.

Staining Protocol

Batch your preparations to avoid over-incubation of samples. Unfixed samples must be acquired within 4 hours after preparation. Samples fixed with 0.2% Guava® fixative, capped, and stored refrigerated are stable for 24 hours.

1. Pipette 10 µL of Human CD4 T Cell Cocktail into each tube.



CAUTION: Place the stock bottle of Human CD4 T Cell Cocktail in the refrigerator or on ice immediately after use. Do not allow the bottle of cocktail to remain at elevated temperatures for an extended time.

2. Add 10 µL of sample to each tube.

For whole blood samples: Blood in the collection tubes should be thoroughly resuspended by gentle agitation for a few minutes before removing an aliquot for sample preparation.

For PBMC samples: Optimal staining is obtained for cell concentrations between 4×10^5 to 2×10^7 cells/mL. Resuspend cells in 1X Assay Buffer BA at appropriate concentrations.

3. Mix the samples thoroughly by pipetting up and down.



CAUTION: Avoid leaving blood to dry on the side of the tubes. This may cause erroneous results.

4. Incubate the samples for 15 minutes at room temperature (18 to 25°C) in the dark.

5. After incubation, add the following:

For whole blood samples: Add 380 µL of Human 1X Lysing Solution directly to each tube to bring the final volume to 400 µL.

For PBMC samples: Add 380 µL of room temperature 1X Assay Buffer BA directly to each tube to bring the final volume to 400 µL.

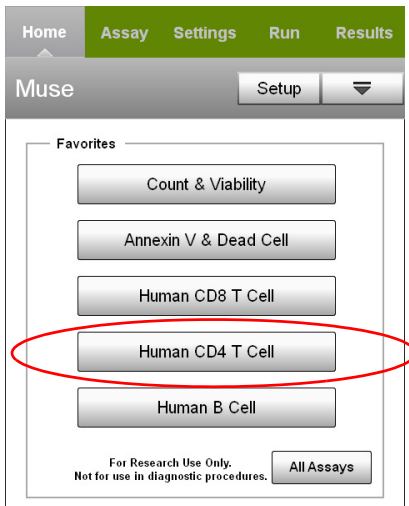
(Optional Protocol with Guava Fixative): If fixation is desired, add 4 µL of Guava Fixative per 400 µL of 1X Lysing Solution or 1X Assay Buffer to a final concentration of 0.2% and use as above.

6. Immediately mix the samples thoroughly by pipetting up and down.
7. **For whole blood samples:** Incubate for a minimum of 15 minutes at room temperature (18 to 25°C) in the dark. After incubation, samples are ready to run on the Guava Muse[®] Cell Analyzer.
For PBMC samples: Run on the Guava Muse Cell Analyzer.

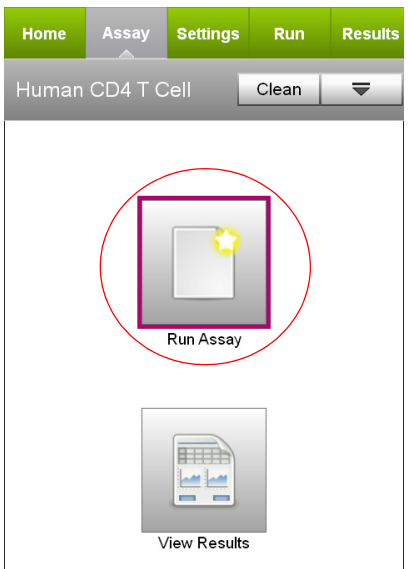
Setup and Acquisition on the Guava[®] Muse[®] Cell Analyzer

Run a System Check prior to performing the assay. For information on Muse[®] System Check, refer to the *Guava Muse Cell Analyzer User's Guide*.

1. Select **Human CD4 T Cell** from the main menu.

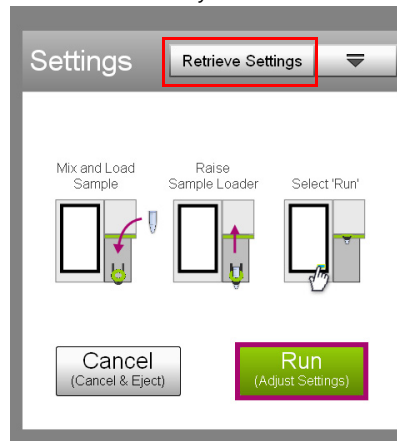


2. Select **Run Assay**.



3. Adjust the instrument settings:

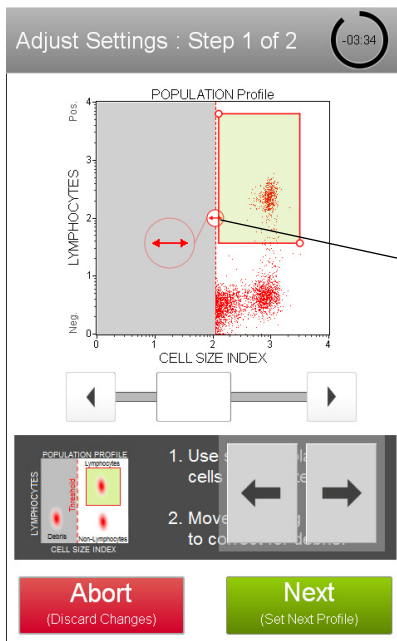
- Load the sample for adjusting the settings and select **Run**. Perform the adjust settings step using a stained sample.
- Or, to retrieve previously saved instrument settings, select **Retrieve Settings**. For more information on retrieving settings, see the *Guava Muse Cell Analyzer User's Guide*.



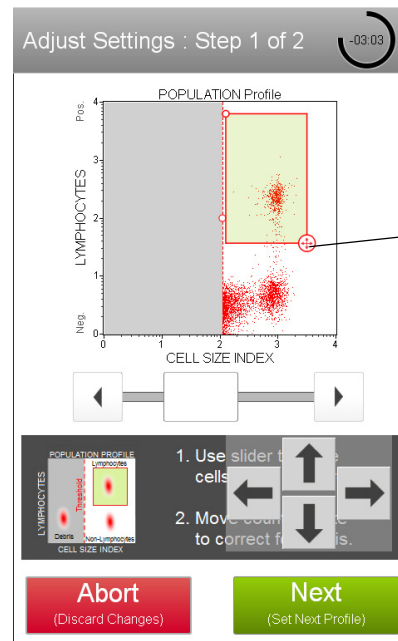
4. Fine tune the settings for the CELL SIZE INDEX vs. LYMPHOCYTE plot, if necessary.

- Adjust the CELL SIZE INDEX slider below the plot to move the lymphocyte population into the green region.
- Drag the threshold to exclude any cellular debris. Touch the threshold and drag to make large changes. The arrow buttons appear after you touch the threshold.

NOTE: If the adjust settings step times out (after 2 minutes), remove the tube and mix well before reloading and continuing. Then, select either **Back** to restart the adjust settings step, or **Next** to accept the settings and continue to the next step.



Touch threshold to move right/left.

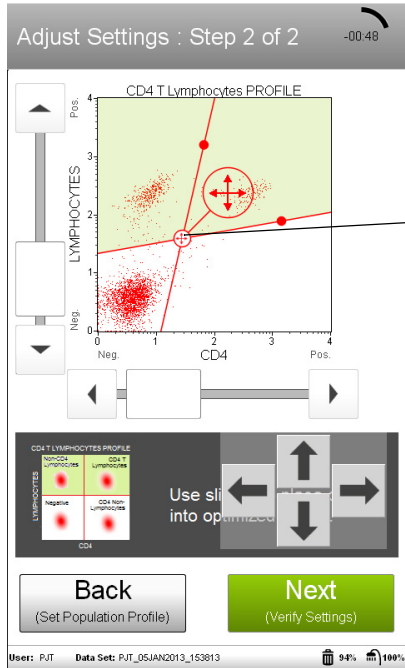


Touch and drag upper-left or lower-right corner to adjust the gate.

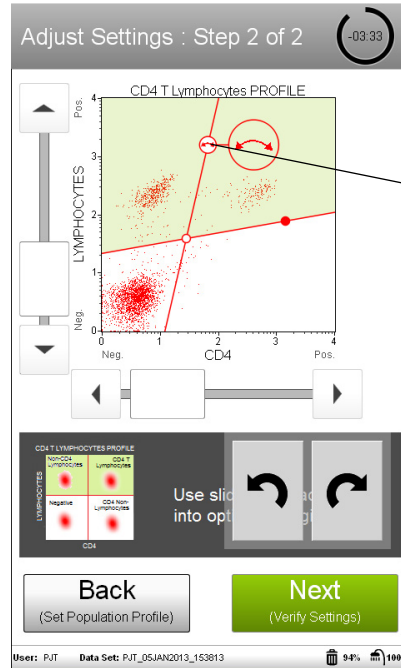
This example data shows typical gate and marker settings. The threshold was raised to remove debris.

5. Select **Next** when you've completed the adjustments.
6. Fine tune the settings for the CD4 vs. LYMPHOCYTES plot, if necessary.

- Adjust the x and y-axis sliders to position the CD4(-) and CD4(+) lymphocytes in the upper-left and upper-right quadrants (green area).
- Adjust the quadrant markers, if necessary. You can move the marker intersection in any direction, as well as adjust the angle of each line. To move the markers as they are, touch the open circle at the intersection and drag the markers to make large changes, or touch the arrow buttons below the plot to make small changes. To adjust the angle of either line, touch the solid circle and drag in an arc, or touch the arrow buttons below the plot.



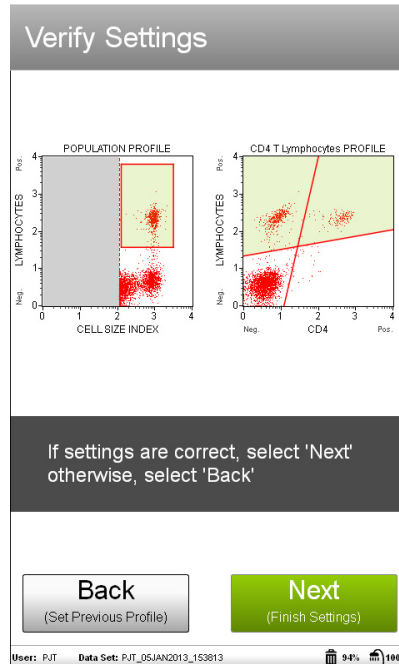
Touch center to move the fixed quadrant markers in any direction.



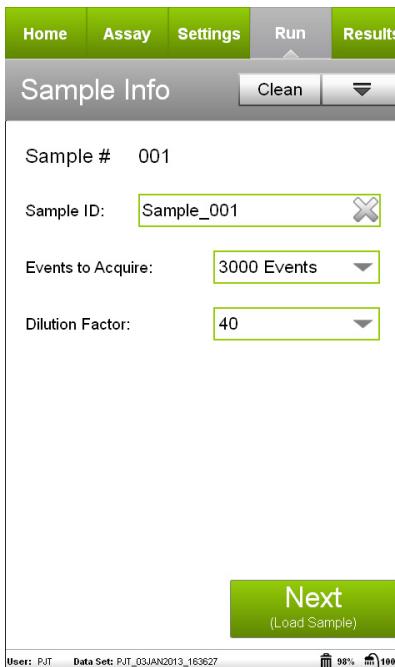
Touch a solid circle on either line (horizontal or vertical) to adjust the angle of the line.

7. Select **Next** when the marker adjustments are complete.

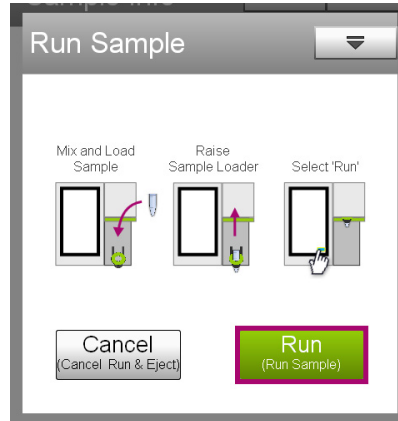
- Verify the settings for the control sample. Select **Back** to make further adjustments. When the settings are correct, select **Next**.



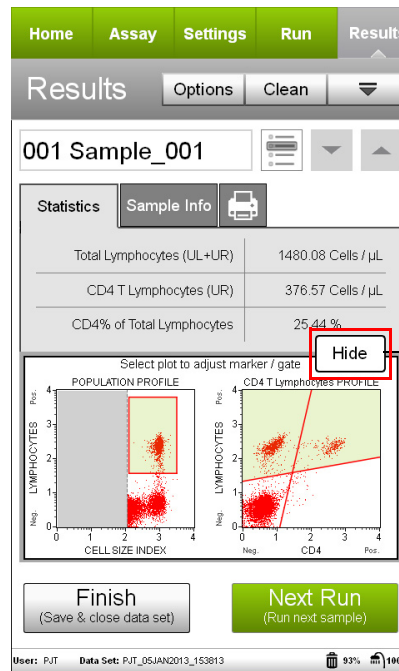
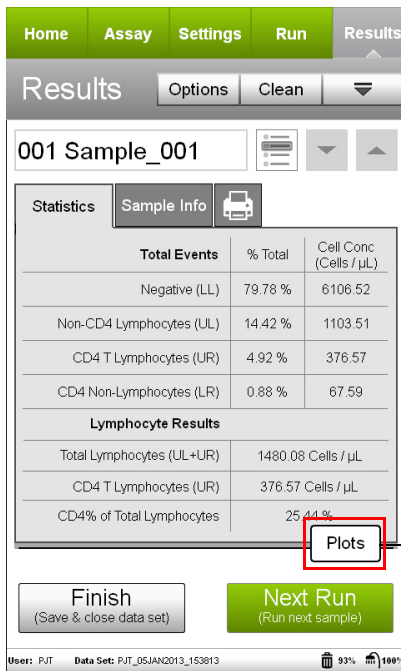
- Enter the sample ID for the first sample by touching the field, then using the keypad to input the ID. Touch **Done** when you finish entering the ID. If necessary, change the **Events to Acquire** and/or **Dilution Factor** by touching the field, then selecting the value from the pop-up menu. Select **Next**.




- Mix the first sample and load it on the instrument. Select **Run** to run the sample.

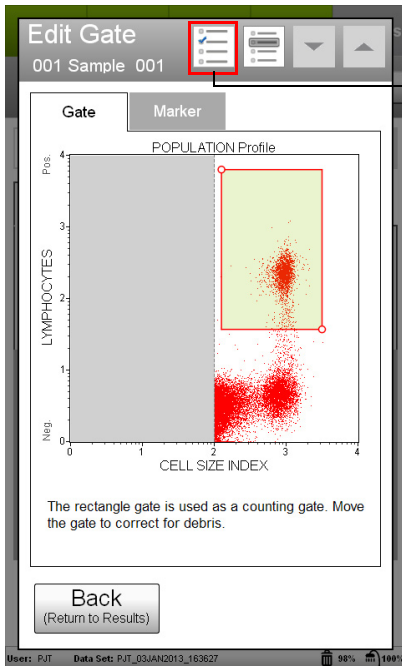


- When acquisition is complete, the results are displayed. Select **Plots** to display dot plots for the sample. You can view or change the sample ID and dilution factor, as well as add annotations for the current sample by selecting the Sample Info tab. To print the results for the current sample select the printer tab.

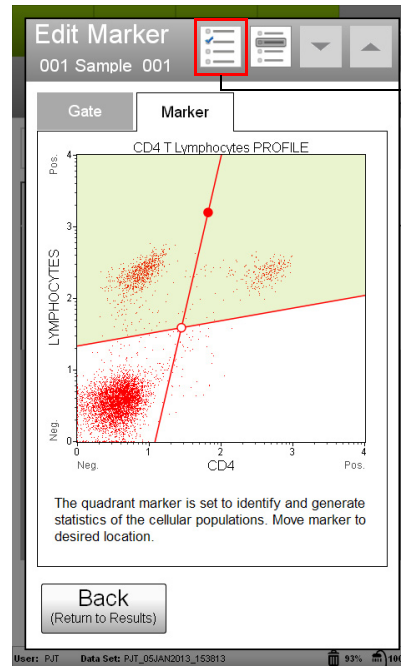


- (Optional) If changes are needed to the gate or markers, touch a plot to enlarge it, then adjust the lymphocyte gate and/or marker as described in steps 4 and 6, respectively. You cannot adjust the cell size threshold after the sample has been acquired.

If you adjust the gate or marker and wish to apply the changes to other samples that you already acquired, select the Apply Changes button () in the title bar. Select the samples you want to apply the changes to or choose **Select All**, then select **Apply**. The sample you originally made changes to must be selected.



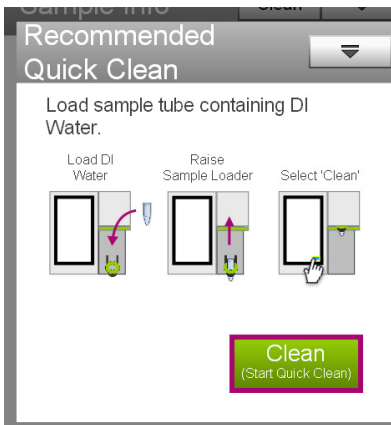
Apply changes



Select to apply changes to other samples.

- If no adjustments are needed, select **Next Run** and repeat steps 9 through 11 for the remaining samples.

NOTE: During the run, a message may appear prompting you to load a tube of DI water for a Quick Clean. Load the water then select **Clean** to perform the Quick Clean. Select **Next** to continue with the run. The frequency of Quick Cleans was set by your system administrator. Your administrator may also have chosen to allow you to skip the Quick Clean when the prompt appears. You can choose to perform additional Quick Cleans at any time during a run by selecting **Clean** in the title bar, then **Quick Clean** from the menu.



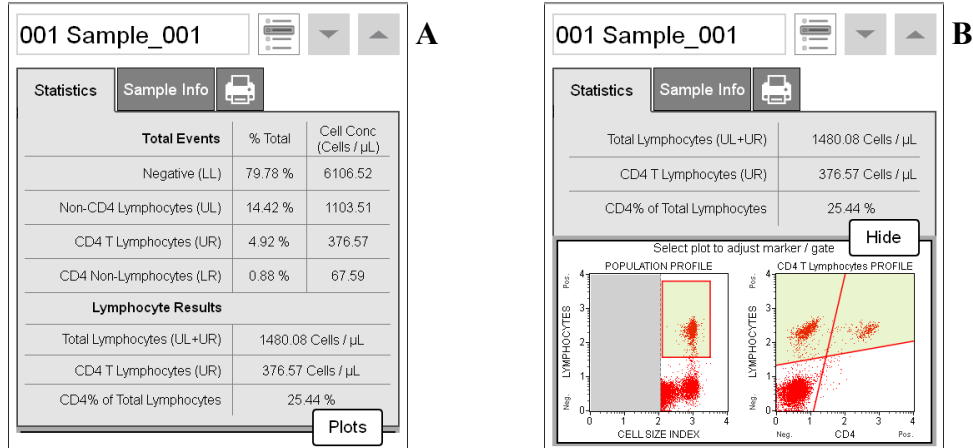
- When you have acquired the last sample, select **Finish**, enter a name for the data set or leave the default, and select **OK**.
- (Optional) Select **Options** in the title bar to rename the data set, export the data set, save the current instrument settings, or view the event log. Refer to the *Guava Muse Cell Analyzer User's Guide* for more information.

Results

Results from each run are stored in a data file, as well as its corresponding spreadsheet (CSV) file. The data file and spreadsheet file contain the following statistics:

- sample number
- sample ID
- percentage of cells in each quadrant
 - upper-left: Non-CD4 lymphocytes [lymphocyte(+) and CD4(-)]
 - upper-right: CD4 T lymphocytes [lymphocyte(+) and CD4(+)]
 - lower-left: Negative cells [lymphocyte(-) and CD4(-)]
 - lower-right: CD4 non-lymphocytes [lymphocyte(-) and CD4(+)]
- concentration and percentage of cells in each quadrant
- CD4 T-lymphocyte concentration, total lymphocyte concentration, and CD4 T-cell percent of lymphocytes
- dilution factor (input value)

Figures A and B. Whole blood was stained with the Muse[®] Human CD4 T Cell Kit and acquired on the Muse Cell Analyzer. Figure A shows summary data, while Figure B shows results displayed with optional dot plots. The statistics show the percentages and the concentrations (cells/ μ L) for the results in each quadrant out of the total events. The Lymphocyte Results show the total lymphocyte concentration, the CD4 T-lymphocyte concentration, and the CD4 cell percentage of lymphocytes. The first plot in Figure B shows Cell Size Index vs. Lymphocytes and a lymphocyte gate, and the second plot shows CD4 vs. Lymphocytes.



Technical Tips

1. All kit reagents, Human CD4 T Cell Cocktail, 1X Assay Buffer BA, and 1X Human Lysing Solution should be brought to room temperature prior to staining.
2. For cellular staining and analysis to be most effective, make sure that test cells have good viability prior to use.

- To clean the instrument after running whole blood, perform a Complete System Clean using two tubes of water instead of Guava® ICF and water. Follow with a second Complete System Clean cycle using Guava ICF, then water, as directed on the screen. See the *Guava Muse® Cell Analyzer User's Guide* for details.

Troubleshooting

Potential Problem	Experimental Suggestions
Acquisition rate decreases dramatically; instrument clogging; too many cells	<ul style="list-style-type: none"> Cell concentration too high: For PBMCs, decrease the cells per microliter by diluting sample to 300-500 cells/μL. The Muse® System gives the most accurate data when the flow rate is less than 500 cells/μL. For whole blood, increase the dilution factor using the Human 1X Lysing Solution to 80. Run a Quick Clean to clean out capillary. This procedure can be performed during or after an assay. This will wash away any material forming within the glass capillary walls.
Too few cells	<ul style="list-style-type: none"> Restain sample using 180 μL of Human 1X Lysing Solution. Ensure dilution factor in the software is set to 20. If using PBMCs, ensure that enough cells were stained as described in "Staining Protocol" on page 5. Restain sample using a more concentrated stock cell suspension.
Background staining and/or non-specific staining of cells	<ul style="list-style-type: none"> Ensure sample was mixed thoroughly prior to acquiring. Ensure dried blood from side of tube was removed prior to staining. Unstained blood will contribute to erroneous results.
Low level of staining	<ul style="list-style-type: none"> Ensure sample was stained for a minimum of 15 minutes followed by complete lysis for 15 minutes. Treated donors may show less separation of cellular populations. Ensure a control sample is run.
Variability in day-to-day and sample-to-sample experiments	<ul style="list-style-type: none"> If the results are inconsistent, check that the samples were well mixed prior to acquisition. Cells may quickly settle in your samples and your results will be inaccurate unless the cells are mixed just prior to acquisition. Ensure that samples are lysed for at least 15 minutes with room temperature Human 1X Lysing Solution. If there appears to be day-to-day variation of the staining pattern, ensure the Muse Cell Analyzer is working properly. Run the System Check procedure using the System Check Kit (Part No. MCH100101) to verify proper instrument function and accuracy.

References

1. Zola H, Swart B., The human leucocyte differentiation antigens (HLDA) workshops: the evolving role of antibodies in research, diagnosis and therapy. *Cell Research*. 2005;12:691-694.
2. Nagel JE, Chrest FJ, Adler WH. Enumeration of T lymphocyte subsets by monoclonal antibodies in young and aged humans. *J of Immunol*. 1981;127:2086-2088.
3. Goetz MB, et al. Relationship between HIV coreceptor tropism and disease progression in persons with untreated chronic HIV infection. *J Acquir Immune Defic Syndrom*. 2009;50:259-266.
4. Huang MC, Patel K, Taub DD, Longo DL, Goetzl EJ. Human CD4-8-T cells are a distinctive immunoregulatory subset. *FASEB J*. 2010;24:2558-2566.
5. Giorgi JV, Hultin LE. Lymphocyte subset alterations and immunophenotyping by flow cytometry in HIV disease. *Clin Immunol Newslett*. 1990;10:55-61.
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Related Kits

- Muse® Human CD8 T Cell Kit - MIM100102
- Muse® Human B Cell Kit - MIM100103
- Muse® Human Lymphocyte CD69 Kit - MIM100104
- Muse® Human Lymphocyte CD25 Kit - MIM100105
- Muse® System Check Kit - MCH100101
- Muse® Count & Viability Kit (40 mL) - MCH100102
- Muse® Annexin V & Dead Cell Kit - MCH100105

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